

# DNase I, GMP-Grade

## Catalog #GMP-DNI-EE001

**Storage Condition** -20°C ± 5°C for 24 months. Avoid repeated freeze/thaw cycles.

**Form** Liquid

**Source** An *E. coli* strain that carries DNase I gene from *Bovine Pancreatic*

**Storage Buffer** 10 mM Tris-HCl (pH7.6), 2 mM CaCl<sub>2</sub>, 50% Glycerol

**Concentration** 4U/μL

**Unit Definition** One unit is the amount of enzyme required to completely degrade 1 μg of pBR322 DNA in 10 minutes at 37 °C.

### Product Contents

- DNase I (RNase-free, 4U/μL)

### Product Description

DNase I is an endonuclease that digests single- and double-stranded DNA to mono- and oligonucleotides containing 5' phosphate and 3'-hydroxylated ends. It is an essential enzyme for the efficient digestion of DNA during RNA purification, suitable for all molecular diagnostics and *in vitro* mRNA synthesis applications.

### Applications

- *In vitro* transcription (IVT)
- Removal of DNA from RNA samples
- DNase I footprinting

### Quality Control Statement

This product has been filed with the FDA Drug Master Files and is assigned DMF #038032. KACTUS manufactures this product according to GMP guidelines and performs stringent quality control testing before release. The production is antibiotic- and animal-free.

### Quality Control Release Criteria

Assay	Criteria
Activity (Plasmids Degradation)	4.0~8.0 kU/mL
Purity (SEC-HPLC)	≥ 95%
Endotoxin	≤ 1.2EU/mL
Residual RNase	Negative
Residual Protease	Negative
Residual Heavy Metal	≤ 10 ppm
Residual Nickel Salt	≤ 10 ppm
Bioburden	≤ 1CFU/10mL
Concentration	0.190 mg/ml±20%

### Protocol for Degradation of DNA Template after *In Vitro* Transcription Reaction

1. After transcription, add 2U of DNase I to a 20μL transcriptional reaction per 1μg DNA template.
2. Incubate at 37°C for 15 minutes.
3. Add EDTA to a final concentration of 5 mM as RNA hydrolyzes during heating with divalent cations in the absence of a chelating reagent.
4. Incubate at 65°C for 10 minutes for heat inactivation of DNase I.

Note: Phenol/chloroform extraction followed by ethanol precipitation of RNA can also be used to remove DNase I.

### Protocol for *In Vitro* Transcription

1. Prepare the following reaction mixture

Reagent	Quantity
5X Transcription Buffer-1 (included with <a href="#">T7 RNA Polymerase</a> )	4μL
CTP/GTP/ATP/UTP (100mM each)	2μL each
<a href="#">Murine RNase Inhibitor</a> (120U/μL)	0.5μL
<a href="#">Pyrophosphatase, Inorganic</a> (0.1U/μL)	1μL
<a href="#">T7 RNA Polymerase</a> (50U/μL)	2μL
Template DNA	1μg
RNase-free Water	Up to 20μL

2. Incubate at 37°C for 1-2 hours.
3. After transcription, add 2U DNase I to digest DNA template for 15 minutes at 37°C.
4. Inactivate DNase I by phenol/chloroform extraction.

### Notes

- Divalent metal ions such as Mg<sup>2+</sup> and Ca<sup>2+</sup> are necessary for DNase I activity, while DNase I is inhibited by monovalent metal ions such as Na<sup>+</sup> and K<sup>+</sup>.